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Co-morbidity of Cystic Fibrosis and Celiac Disease in an Indian Female Child

R K Gupta, Dhan Raj Bagri, Akash Soni, Isha Gupta

Department of Pediatrics, SMS Medical College, Jaipur-302004, India.

Corresponding Author: Dr Dhan Raj Bagri Email: meena.drdhanraj6@gmail.com

Cystic fibrosis (CF; OMIM: 219700), is an autosomal recessive disease caused by the homozygous or compound heterozygous mutations of the cystic fibrosis transmembrane conductance regulator (CFTR) gene (OMIM*602421) encoding the CFTR protein. It is the most common lethal genetic disease among Caucasians, with a reported birth prevalence of 1:2000-1:4000 live births. The CFTR dysfunction affects epithelial cells, compromising innate and adaptive immune systems both in terms of quality and quantity of response resulting in

immunological imbalance; which provides the basis for developing autoimmune diseases (AID) as comorbid conditions. In CF, the most investigated AID is celiac disease.¹

The risk of the co-morbidity of CF and CD has also been suggested previously. The incidence of the two conditions occurring in the same patient has been estimated to be 1 in 2,000,000-1:5,900,000. Several case– control studies reporting a wide range (1.2-2.13%) of proven CD incidence in European CF patients have been published in the last decade².

We report a case in which the female child was diagnosed with this co-morbidity at 14 years of age. We conclude that despite the different pathophysiology, both CF and CD cause malabsorption; and pose a diagnostic dilemma for pediatricians in distinguishing between CFrelated or potentially CD-related GI symptoms. A better understanding of the comorbidity of both diseases is needed for early diagnosis and management.

Case Report

A 14 Years old female child, known patient of celiac disease for the last 10 years; on a gluten-free diet, presented with complaints of recurrent respiratory infections, undocumented fever in the last year and cough in the last 6 months. Fever had a diurnal variation with evening rise of temperature and was aggravated in the last 20 days. Cough was mucoid, greenish-colored, non-blood stained, and associated with respiratory difficulty and fever. She also has a history of oily stools, constipation and failure to gain weight. The patient kept on taking medications from various pediatricians with symptomatic relief only.

She was hospitalized at 3 years of age for cough and respiratory distress at 4 years of age with complaints of constipation and surgery for bowel obstruction, and at 7 years of age for cough, fever and blood transfusion. Her mother's younger sister was also diagnosed with celiac disease. Her antenatal course was uneventful but there was history of meconium-stained liquor. She was given mixed feeding and weaning was started at 7 months. She is incompletely immunized. Currently, she is taking milk, makhane, cornflakes, 2 chapatis of celiac flour, vegetables, dal and tea.

On examination, her height was 138 cm which was < 3rd centile and her weight was 23 kg which was also < 3rd centile for age. Her body mass index (BMI) was also < 3rd centile. Digital clubbing on both hands and right nasal polyp with inferior turbinate hypertrophy was noted, and nasal mucosa was pink. The tongue was pink and dental hygiene was adequate. Respiratory system examination revealed symmetric bilateral chest movements and expansion with no visible scars or swelling. Trachea was midline. Air entry was equal with resonant bilateral lungs. Coarse crepitations were present on both sides. Abdomen was scaphoid and all quadrants moved equally with respiration. A scar of 6x1 cm was present on the right lumbar region. No visible veins were noted. The abdomen was soft and bowel sounds were audible. No hepatosplenomegaly was noted.

Her complete blood counts were normal with slight leukocytosis of $11.12 \times 10^3/\mu$ l, with normal differential counts. Hemoglobin was 12.9 gm/dl with normal MCV, MCH and MCHC and normal platelet counts. Investigations have been summarized in **Table-1** and **Figure 1,2**. Sputum for AFB and CBNAAT were negative. The diagnosis of celiac disease was confirmed by upper GI endoscopy and duodenal biopsy (modified Marsh Type 2). Positive HLA DQ2.5 endorsed the diagnosis.

CECT Scan of the thorax revealed multiple centrilobular nodules (tree-in bud pattern) in the anterior segment of the bilateral upper lobe, the lateral segment of the right middle lobe, the anterior-basal segment, the lateral-basal segment of the right lower lobe, lingula of left upper lobe, the superior-basal, antero-medial-basal

LH	0.62 m IU/ml		Follicular phase -1.2-12.5 m IU/ml Ovulatory period-12-82 mlU/ml Luteal phase-0.4-19 mlU/ml Menopause -14-48 mlU/ml		
FSH	2.01 m IU/ml		Follicular phase-3.2-15 m IU/ml Ovulatory phase-7.5-20 mlU/ml Luteal phase-1.3-11 mlU/ml Menopause-36-138 mlU/mI		
Free T4	13.64 pg/ml		8.5-22.50		
TSH	3.58 mIU/ml		0.30-4.50		
Free T3	3.05 pg/ml		1.21-4.30		
Anti TPO Antibody	0.19		0.90-1.10		
Insulin	D3 1.52 µ IU/ml	D7 5.72 μ IU/ml	4.03-23.46		
C-peptide	0.53 ng/ml		0.80-4.20		
HbA1C	6.3%		<6		
Prolactin	9.72 ng/ml		3.10-23.03		
IgA tTG	1.24 AU/ ml		< 8.0 AU/ml		
SGOT	43.89 U/L		< 35.00		
SGPT	15.66 U/L		<40.00		
Amylase	71.63 U/L		<140.00		
Lipase	28.63 U/L		<60.00		

Table 1: Blood investigations

Table 2: HLA Typing for Celiac Disease

Allele Name	Result
DRB1*03DQA1*05:01-DQB1*02:01	Positive
DRB1*07-DQAI*02:01-DQB1*02:02	Negative
DRBI*II-DQA1*05:05DQB1*03:01	Negative
DRB1*04-DQA1*03:01-DQB1*03:02	Negative



Figure 1: X-Ray Wrist

and lateral basal segment of left lower lobe. Patchy ground glass haziness was noted in the anterior segment of the left upper lobe and the posterior basal segment of the left lower lobe. Bronchial wall thickening was noted in the bilateral lung parenchyma. No evidence of enlarged lymph nodes was seen in the mediastinum and hilar regions. Overall findings were suggestive of bronchial wall thickening with centrilobular nodules and ground glass haziness consistent with infective bronchiolitis.

MRI of abdomen and pelvis with gadolinium (Tech: SE TI & FSE -T2 weighted axial & Coronal images of the upper abdomen along with axial T 1, T2 and coronal FSIR & sag T2W images of the lower abdomen); revealed a fibrotic pancreas and both ovaries had antral follicles arranged peripherally giving a necklace-like appearance, with thickened stroma suggestive of polycystic ovarian disease (PCOD). The uterus was very small in size. Rest of the intraabdominal organis were normal (**Figure-3**).

Tests for the exocrine pancreatic function showed decreased insulin and C-peptide levels but no hyperglycemia was documented or reported. Immunoreactive trypsinogen for the immunoreactive trypsinogen was reported as negative.

Whole Exome sequencing detected likely



Figure 2: Chest X Ray

pathogenic variant causative of the reported phenotype (Table 3). A homozygous start loss variation (that alters the ATG start codon and consequently affects its translation) in exon 1 of the CFTR gene (chr7: g.117480096T>C; Depth:104x) (p.Met 17; ENST0000000-3084,11) was detected. The observed variant has previously been reported as p. MIT in patients with cystic fibrosis. The p. Met1 (suspected) variant has not been reported in the 1000 genomes and genome databases and had a minor allele frequency of 0.015% in concerned internal database. The in-silico prediction of the variant damages by SIFT, LRT, and mutation Taster 2. The reference codon is conserved across species. The child was given protocol management with a gluten-free diet and nutritional counselling. She got improved and was discharged with the advice of chest physiotherapy and education for symptoms of pulmonary exacerbation.

Discussion

Cystic fibrosis is characterized by a triad of chronic obstructive pulmonary disease, exocrine pancreatic insufficiency, and elevation of sodium and chloride concentration in sweat. Immune dysregulation in

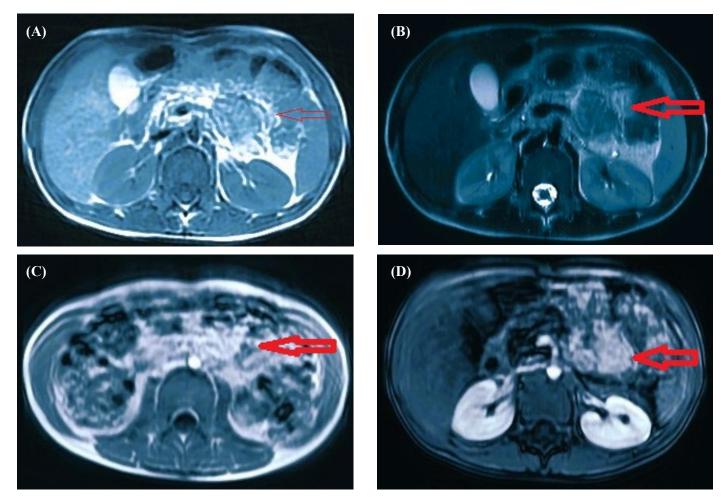


Figure 3 (A): MRI Abdomen and Pelvis Axial T1 W image showing atrophy of distal body and tail of pancreas and increased T1 intensity of head and proximal body of pancreas, (B): MRI Abdomen and Pelvis Axial fat sat T2 image showing suppression of fat in remaining head and proximal body of pancreas, s/o macroscopic fat replacement, (C): MRI Abdomen and Pelvis- Axial in phase image showing atrophy of distal body and tail of pancreas, (D): MRI Abdomen and Pelvis- Axial out phase image showing fatty replacement of remaining head and proximal body of pancreas showing suppression on chemical shift imaging, suggestive of microscopic fat.

Table 3: Whole exome sequencing

Gene (Transcript)	Location	Variant	Zygosity	Disease (OMIM)	Inheritance	Classification
CFTR (+) (ENST0000000-	Exon 1	C2T>C	Homozygous	Cystic Fibrosis	Autosomal	Likely Pathogenic
3084,11)		(p.Met1?)			Recessive	

CF is hypothesized to be due to lipopolysaccharide hypersensitivity of alveolar macrophages; enhanced signal transduction via NF- κ B and MAPK pathways; altered LPS-induced metabolic pathways; impaired apoptosis of granulocytes; and an increased release of various proinflammatory cytokines, including IL-17 and IL-6, etc.²⁻⁵.

The child in our case report was on a glutenfree diet and was well conversant with the importance of a GFD. Despite a better understanding of the course, nature and prognosis of disease; the child didn't grow well and suffered recurrent illnesses and could only be diagnosed to have co-morbidity of cystic fibrosis at a later age. The heavy burden of undigested gluten proteins in the intestine due to exocrine pancreatic insufficiency, intestinal mucosal damage due to malnutrition and an immunological response against wheat gluten may explain her condition.

Conclusion

Clinical symptoms and signs of CD are difficult to differentiate from CF with malabsorption, and we, therefore, suggest that serological screening for CD should be included in the diagnostic work-up of CF patients older than nine months and patients with persistent gastrointestinal symptoms. Such case reports suggest the importance of screening for CD in CF and vice versa. Further studies should be undertaken for the estimation of the prevalence of CF in Indian CD children. The clinical phenotype of CF with and without CD should be described in detail to define those who might benefit from CD screening and, later, from the gluten-free diet.

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Hepatitis C Virus-Autoimmune Hepatitis (HCV-AIH) Overlap Syndrome:ATreatmentDilemma

Deepanshu Khanna, Premashis Kar

Department of Gastroenterology, Max Superspeciality Hospital, Vaishali, Ghaziabad, U.P, India.

Corresponding Author: Dr Deepanshu Khanna Email: drdeepanshukhanna@gmail.com

HCV-AIH overlap syndrome is a rare condition that poses challenges in detection and treatment. The diagnosis is typically established when the patient exhibits simultaneous clinical, serologic, and histologic signs of both diseases. However, due to possible similarities between the histologic and serologic markers of HCV and AIH, use of precise diagnostic standards is crucial to avoid unnecessary medical interventions.

Despite being specific for AIH, autoantibodies such as ANA, anti-SMA, and anti-LKM-1 are also commonly found in HCV patients. Portal lymphoid aggregates or reactive lymphoid follicles, with or without lobular activity are the histological features of HCV¹. The International Autoimmune Hepatitis Group proposed that interface hepatitis with predominant plasma cell infiltrate, emperipolesis, and hepatocellular rosette formation are the hallmark morphologic features of AIH. HCV and AIH may occasionally exhibit identical histologic findings, when they are in their chronic forms (chronic hepatitis with associated fibrosis).

Currently, there are no accepted guidelines for the care of HCV-AIH patients. In the past, it was known that interferon-based treatment for HCV could lead to development of various autoimmune diseases, including AIH. AIH could manifest not only during or immediately after the treatment but also long after the therapy was stopped. Although immunosuppressive drugs have the potential to make HCV infection worse by promoting viral replication, corticosteroids have shown to be a modestly effective first-line treatment for HCV-AIH patients².